

DEFICIENCY OF COENZYME  $Q_{10}$  IN HYPERTENSIVE RATS  
AND REDUCTION OF DEFICIENCY BY TREATMENT WITH COENZYME  $Q_{10}$

by

Yoshifumi Iwamoto, Toru Yamagami, and Karl Folkers

Institute for Biomedical Research  
The University of Texas at Austin  
Austin, Texas 78712

and

C. Gunnar Blomqvist

Department of Internal Medicine  
Laboratory for Cardiopulmonary Research  
University of Texas  
Health Science Center at Dallas  
Dallas, Texas 75235

Received April 13, 1974

SUMMARY

Rats which were unilaterally nephrectomized and treated with deoxycorticosterone acetate and saline developed hypertension, and had an increased heart weight. Treatment of these rats with coenzyme  $Q_{10}$  effected a lower level of hypertension and normalized the heart weight. The specific activities of the succinate dehydrogenase- $CoQ_{10}$  reductase of leucocytes from these hypertensive rats showed an increase ( $P < 0.05$ ) in the deficiency of  $CoQ_{10}$ -enzyme activity; treatment with  $CoQ_{10}$  lowered ( $P < 0.05$ ) the activity to that of the normal group. Leucocytes are convenient and sensitive for monitoring  $CoQ_{10}$ -enzymes. A deficiency of  $CoQ_{10}$  in hypertension is undesirable for effective bioenergetics and might be corrected by therapy with  $CoQ_{10}$ .

INTRODUCTION

Yamagami *et al.* (1) reported upon a deficiency of coenzyme  $Q_{10}$  in leucocytes from patients having hypertension. They found that the specific activity of the succinate dehydrogenase-coenzyme  $Q_{10}$  reductase in mitochondria from leucocytes from certain patients with essential hypertension was not only lower ( $P < 0.001$ ), but was deficient ( $P < 0.001$ ) in comparison with healthy individuals with good nutrition. They summarized information supporting the suggestion that a deficiency of coenzyme  $Q_{10}$  may be of some clinical significance in human hypertension.

Igarashi *et al.* (2) and Yamagami *et al.* (3) observed a partial but not a total reduction of the experimental hypertension in rats, which is induced by

deoxycorticosterone acetate and saline, when such rats were treated with coenzyme Q<sub>10</sub>. Igarashi *et al.* (2) investigated the content of sodium and potassium in the plasma and in the tissues of these hypertensive rats, and found that the Na/K ratio was higher in the hypertensive than in the normal rats, and that the Coenzyme Q. 182.

ratio decreased with treatment with coenzyme Q<sub>10</sub>. Yamagami *et al.* (3) observed that a component of the "antihypertensive" action of coenzyme Q<sub>10</sub> might be based on the decrease of the hypersensitivity of the peripheral vessels to norepinephrine.

The specific activities of the succinate dehydrogenase-coenzyme Q<sub>10</sub> reductase in mitochondrial preparations from the leucocytes, liver, kidney, and heart from these hypertensive rats have been determined, and after such rats were treated with coenzyme Q<sub>10</sub>.

#### MATERIALS AND METHODS

Thirty-five rats, weighing 180-200 g, were divided into three groups. Twelve rats constituted a normal control group, and were fed a customary laboratory diet. Another group of 12 rats were unilaterally nephrectomized, and were orally treated with 0.9% saline, and 25 mg/kg of deoxycorticosterone acetate (DOCA) by intramuscular injection once a week to induce hypertension. Hypertension was similarly induced into a third group of 11 rats which were orally treated with coenzyme Q<sub>10</sub> dissolved in corn oil. The dose of coenzyme Q<sub>10</sub> was 50 mg/kg during the first 4 weeks and then the dose was increased to 100 mg/kg for the fifth and sixth weeks. The systolic blood pressure was recorded every week as described (3). The rats were sacrificed after 6 weeks by decapitation, and whole blood, hearts, kidneys, and livers were removed for enzyme assays.

The blood samples from the rats were processed and assayed as described by Nakamura *et al.* (4) for the determination of the specific activities of the succinate dehydrogenase-coenzyme Q reductase, and the tissue of the hearts, kidneys, and livers were processed essentially as described by Littarru *et al.* (5) for the enzyme assay.

#### RESULTS

Yamagami *et al.* (3) described studies on the reduction by coenzyme Q<sub>10</sub> of experimental hypertension in rats which is induced by deoxycorticosterone and saline. They also related their studies to the literature and the citations and relationships are not herein repeated. They showed the partial but not complete control by coenzyme Q<sub>10</sub> of hypertension in this animal model. The systolic blood pressure was elevated in both the untreated and CoQ<sub>10</sub>-treated groups ( $P < 0.001$ ) in comparison with that of the normal control group. This induced hypertension

was partially lowered ( $P < 0.05$ ) by the treatment with coenzyme  $Q_{10}$  and confirmed, in principle, data reported by Igarashi *et al.* (2).

The mean body weights in the normal control group was about 321 g, about 262 g ( $P < 0.001$ ) in the untreated hypertensive group, and about 266 g ( $P < 0.001$ ) in the  $CoQ_{10}$ -treated group. It was particularly noted that the mean weights of the hearts of the non-treated hypertensive group was greater ( $P < 0.05$ ) than that of the normal control group, and that treating these hypertensive rats with coenzyme  $Q_{10}$  apparently prevented the enlargement of the heart and its increase in weight.

The data on the specific activities and the percent deficiencies of enzyme activities of the succinate dehydrogenase-coenzyme Q reductase ( $CoQ_{10}$ -enzyme) for the leucocytes, livers, kidneys, and hearts of the three groups of rats which were sacrificed after the test period of 6 weeks are in Table 1.

The percent deficiency of the coenzyme  $Q_{10}$ -enzyme of the livers of the  $CoQ$ -treated hypertensive rats was lower ( $P < 0.01$ ) than that of the normal rats and that of the untreated controls. The livers of the normal control rats appeared deficient in  $CoQ_{10}$ , and such deficiency was largely corrected by the administration of  $CoQ_{10}$ . Similarly, the kidneys of the normal control rats also appeared

Table 1. DATA ON SUCCINATE DEHYDROGENASE-COQ REDUCTASE OF RAT TISSUES

Groups	Specific Activity	Specific Activity with $CoQ_3$	Activation Coefficient	% Deficiency $CoQ_{10}$ -Enzyme Activity
<b>BLOOD</b>				
Normal	1.87+0.30	2.59+0.24	42.17+7.68	27.00+3.60
Hypertensive	2.70+0.30	4.63+0.46	76.00+8.63	41.42+3.19
Hypertensive- $CoQ_{10}$ Treated	3.09+0.34	4.51+0.55	47.82+9.70	29.09+5.02
<b>LIVER</b>				
Normal	38.47+2.99	73.63+ 4.78	95.25+9.50	47.42+2.86
Hypertensive	42.78+5.59	85.81+14.99	95.42+6.64	48.25+1.59
Hypertensive- $CoQ_{10}$ Treated	71.17+8.02	102.34+ 8.01	51.82+8.49	32.18+3.59
<b>KIDNEY</b>				
Normal	53.08+3.72	94.08+6.51	78.08+4.97	43.42+1.92
Hypertensive	78.05+7.29	124.68+6.41	66.08+8.12	37.83+3.62
Hypertensive- $CoQ_{10}$ Treated	93.64+7.76	138.86+9.43	50.91+4.33	33.18+1.93
<b>HEART</b>				
Normal	107.79+11.46	145.43+11.46	39.42+6.89	26.08+4.03
Hypertensive	133.21+ 9.24	195.49+14.09	49.25+4.34	32.50+1.94
Hypertensive- $CoQ_{10}$ Treated	148.18+ 8.15	195.86+ 8.34	35.36+4.78	25.18+3.01

Values are Mean + S.E.

deficient in coenzyme  $Q_{10}$  and which was substantially absent in the  $CoQ_{10}$ -treated rats. The expression "normal" rat is relative for specific reasons; it has been previously observed (6) that so-called normal animals which are maintained on ordinarily purchased diets are not necessarily normal in respect to saturation with  $CoQ$  of  $CoQ_{10}$ -enzymes in the tissue, and probably in other nutritional aspects. As the levels of coenzyme  $Q_{10}$  varies in those tissues of rats which are important to the pathogenesis of the hypertension, the degree of hypertension to be elicited when such "normal" rats are treated with DOCA-saline may be expected to vary.

In the leucocytes of the blood of the untreated hypertensive rats, the greatest increase in the percent of deficiency of  $CoQ_{10}$ -enzyme activity was observed. The percent deficiencies in the kidneys and livers showed no difference between the normal rats and the untreated hypertensive rats, but a significant decrease ( $P < 0.01$ ) in comparison with normal rats when  $CoQ_{10}$  was administered to the hypertensive rats.

The actual level of  $CoQ_{10}$ -enzyme activity is the highest in heart tissue and the lowest in the leucocyte preparations from the blood. Regardless of the absence or presence of coenzyme  $Q_3$  in the differential assay, and for all three groups of rats, these relative levels are compatible with previous studies and reflect again the importance of bioenergetics to the function of the heart.

It is meaningful to appraise the data on the percent deficiency of  $CoQ_{10}$ -enzyme activity, because this criterion is directly correlated with a deficiency of coenzyme  $Q_{10}$  in the tissue from which the mitochondrial preparation was made.

The leucocyte preparations from the hypertensive rats showed a significant deficiency ( $P < 0.05$ ) in  $CoQ_{10}$ -enzyme activity in comparison with that of the normal control group. The hypertensive rats which were treated with coenzyme  $Q_{10}$  did not show a significant deficiency as was observed for the normal control group. Percent deficiencies of 25-30% have been observed for rats maintained on ordinary laboratory diets with no special precautions against oxidation of unsaturated lipids in the diet.

## DISCUSSION

The data in Table 1 were obtained using rats with an ordinary dietary background and from a normal supplier where there was no effort before or after the rats were obtained to have truly normal saturation with coenzyme  $Q_{10}$  of coenzyme  $Q_{10}$ -enzymes within mitochondria and the Golgi apparatus. For example, treatment of hypertensive rats with coenzyme  $Q_{10}$  reduced the deficiencies of activity of the  $CoQ_{10}$ -enzyme for the livers and hearts to a significant degree ( $P < 0.01$ ). However, the hypertensive state did not lead to an increase in the deficiency

of activity in the liver, kidney and heart in comparison with these organs of the normal control group.

The leucocytes of the blood were the most sensitive and revealed a significant increase ( $P < 0.05$ ) in deficiency of CoQ<sub>10</sub>-enzyme activity. The leucocytes of the hypertensive rats which had been treated with coenzyme Q<sub>10</sub> revealed a significant reduction ( $P < 0.05$ ) of the deficiency. It is fortunate that the leucocytes show this sensitivity to the hypertensive state in response to treatment with coenzyme Q<sub>10</sub>, because leucocyte levels can be monitored without sacrifice of the animal.

Whether there is a correlation between the levels of saturation with coenzyme Q<sub>10</sub> in its enzymes in leucocytes and the lowering of hypertension by treatment with coenzyme Q<sub>10</sub> is not clear, but it seems reasonable.

Perhaps the hypertensive state increases the need for coenzyme Q<sub>10</sub> which is not fulfilled by increased biosynthesis but can be provided through treatment with coenzyme Q<sub>10</sub>.

A new methodology to detect and measure vitamin deficiencies in mammalian tissues has been reviewed by Folkers *et al.* (7). The methodology is based upon the enzyme activity of a coenzyme-apoenzyme complex in which the vitamin or its coenzyme form is indispensable to the activity of the enzyme complex. The principle of this assay was formulated by Folkers (8) as follows -- "The specific activity of a coenzyme-apoenzyme system is differentially assayed in the absence and in the presence of added coenzyme. A significant increase in the specific activity of the enzyme system in the presence of added coenzyme indicates a deficiency of the coenzyme at its site or of the vitamin in the tissue".

Brin *et al.* (9) first utilized this principle to determine deficiencies of thiamin. Glatzle *et al.* (10) similarly studied deficiencies of riboflavin in geriatric patients. Raika and Sauberlich (11) and Krishnaswamy (12) assayed for deficiencies of vitamin B<sub>6</sub> by this principle.

The extension of this principle to a new enzymatic assay for human deficiencies of coenzyme Q<sub>10</sub> was detailed by Nakamura *et al.* (13) and this same assay may be used to seek and measure deficiencies of coenzyme Q<sub>10</sub> in experimental animal tissues including tissues of the hypertensive rats. Generally, the dual assay is performed on the mitochondrial succinate dehydrogenase-coenzyme Q<sub>10</sub> reductase, but it can also be applied to DPNH-cytochrome c reductase.

Data showing certain deficiencies of coenzyme Q<sub>10</sub> in biopsies of human hearts were reported by Littarru *et al.* (14). The specific activity of the succinate-dehydrogenase-coenzyme Q<sub>10</sub> reductase of the ventricle wall of the human heart was about 123 nmoles/mg/min. Littarru *et al.* (5) found about the same level (S.A. 141) of enzyme activity for the heart tissue of the rabbit. The specific activity of this CoQ<sub>10</sub>-enzyme of the normal rats (Table 1) was 108 and

in the presence of coenzyme  $Q_3$ , the specific activity was 145. For blood, the specific activity of this  $CoQ_{10}$ -enzyme in the leucocytes was found by Nakamura *et al.* (13) to be 1.56 for healthy persons and that of the normal rats (Table 1) was 1.87. It is apparent that the level of specific activity of this  $CoQ_{10}$ -enzyme is approximately the same in humans, rabbits and rats.

It can be suggested that an increased deficiency of coenzyme  $Q_{10}$  in the hypertensive state is an undesirable condition for effective bioenergetics and particularly as required for ion transport including sodium. Correction of a deficiency of coenzyme  $Q_{10}$  would surely be a desirable contribution to the control of hypertension.

#### ACKNOWLEDGMENT

Appreciation is expressed to the Hartford Foundation and The Robert A. Welch Foundation for their respective support. Mrs. Chung Hee Kim provided very skillful technical assistance.

#### REFERENCES

1. Yamagami, T., Iwamoto, Y., Folkers, K., and Blomqvist, C.G., International J. Vit. and Nutr. Res. (in press).
2. Igarashi, T., Tanabe, Y., Nakajima, Y., Kobayashi, M., Tanaka, M., and Ohtake, S., Folia Pharmacol. Japon., **68**, 460 (1972).
3. Yamagami, T., Iwamoto, Y., Folkers, K., and Blomqvist, C.G., International J. Vit. and Nutr. Res. (in press).
4. Nakamura, R., Littarru, G.P., and Folkers, K., International J. Vit. and Nutr. Res., **43**, No. 3, 296 (1973).
5. Littarru, G.P., Jones, D., Scholler, J., and Folkers, K., International J. Vit. and Nutr. Res., **42**, 127 (1972).
6. Scholler, J., Jones, D., Littarru, G.P., and Folkers, K., Biochem. Biophys. Res. Commun., **41**, No. 5, 1298 (1970).
7. Folkers, K., Littarru, G.P., Nakamura, R., and Scholler, J., International J. Vit. and Nutr. Res., **42**, 139 (1966).
8. Folkers, K., American Journal of Clinical Nutrition (in press).
9. Brin, M., Tai, M., Ostashever, A.S., and Kalinsky, H., J. Nutr., **71**, 273 (1960).
10. Glazle, D., Körner, W.F., Christeller, S., and Wiss, O., International J. Vit. and Nutr. Res., **40**, 166 (1970).
11. Raika, N. and Sauberlich, H.E., Am. J. Clin. Nutr., **15**, 67 (1964).
12. Krishnaswamy, K., International J. Vit. and Nutr. Res., **41**, 240 (1971).
13. Nakamura, R., Littarru, G.P., and Folkers, K., International J. Vit. and Nutr. Res., **43**, No. 4, 526 (1973).
14. Littarru, G.P., Ho, L., and Folkers, K., International J. Vit. and Nutr. Res., **42**, 413 (1972).